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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁷ : C12N 15/00, 15/12, 15/13, 15/81, C07K 16/00, G01N 33/50, A61K 39/395	A1	(11) International Publication Number: WO 00/23579 (43) International Publication Date: 27 April 2000 (27.04.00)
(21) International Application Number: PCT/US99/24677 (22) International Filing Date: 21 October 1999 (21.10.99) (30) Priority Data: 60/105,259 22 October 1998 (22.10.98) US (71) Applicant: THE REGENTS OF THE UNIVERSITY OF CALIFORNIA [US/US]; 1111 Franklin Street, 12th floor, Oakland, CA 94607-5200 (US). (72) Inventors: CHOUDARY, Prabhakara, V.; Apartment #4, 422 Russell Park, Davis, CA 95616 (US). OGUNJIMI, Abiodun, A.; 6 Edmond Crescent, PMB 2013, Yaba, Lagos (NE). CHANDLER, John, M.; 2648 Lafayette Drive, Davis, CA 95616 (US). (74) Agent: BERLINER, Robert; Fulbright & Jaworski L.L.P., 29th floor, 865 S. Figueroa Street, Los Angeles, CA 90017-2576 (US).		(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i>
(54) Title: FUNCTIONALLY ASSEMBLED ANTIGEN-SPECIFIC INTACT RECOMBINANT ANTIBODY AND A METHOD FOR PRODUCTION THEREOF (57) Abstract Functionally assembled antigen-specific intact recombinant monoclonal antibody produced by transformation of the methylotropic yeast, <i>P. pastoris</i> with mouse/human immunoglobulin genes encoding heavy and light chains. A method for production of the intact monoclonal antibodies, a recombinant yeast expression vector and the antibody-specific mRNA synthesis. A process for a large-scale production of the functionally assembled intact recombinant antibody.		

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FUNCTIONALLY ASSEMBLED ANTIGEN-SPECIFIC INTACT
RECOMBINANT ANTIBODY AND A METHOD FOR PRODUCTION THEREOF

This application is based on and claims priority of
5 the Provisional Application Ser. No. 60/105,259 filed on
October 22, 1998.

BACKGROUND OF THE INVENTION

Field of Invention

This invention concerns functionally assembled
10 antigen-specific intact recombinant monoclonal antibody
produced by transformation of the methylotropic yeast,
Pichia pastoris transformed with immunoglobulin (Ig) genes.
In particular, this invention concerns production of
immunologically active antigen-specific intact recombinant
15 mammalian, including human, antibody, transformed with
immunoglobulin genes. The invention also concerns a method
and process for production of the intact monoclonal
antibody, a recombinant yeast expression vector and the
antigen-specific antibody synthesis. The invention further
20 concerns a method for large-scale production of the
functionally assembled intact recombinant mammalian,
including human, antibody.

BACKGROUND AND RELATED DISCLOSURES

Recombinant DNA technology has facilitated
25 humanization of murine monoclonal antibodies (Ann. Allergy
Asthma Immunol., 81:105, (1998)) and heterologous production
of antibody fragments (Res. Immunol., 149:587, (1998)). As
a result, over the past 15 years, numerous antibody
fragments, such as Fab, 30 FV, scFv, or diabodies, have
30 been produced in bacterial hosts (Curr. Opinion Microbiol.,
5:256, (1993)). Prokaryotes, however, are incapable of
producing complex multimeric glycoproteins, such as intact
antibodies, which require posttranslational modifications
in a functionally assembled form. Prokaryotes also tend to
35 accumulate over-expressed recombinant proteins as insoluble

inclusion bodies, necessitating additional denaturation-
renaturation steps for recovering recombinant proteins.
These steps often impair the biological function of these
recombinant proteins. As an alternative, several
5 eukaryotic hosts have been evaluated for ability to produce
functionally assembled intact antibodies (New Frontiers in
Agrochemical Immunology, 171-186, D.A. Kurtz et al, AOAC
International, Arlington, Virginia (1995)).

Mammalian cell lines have been previously investigated
10 with some degree of success as hosts for recombinant
antibody production. However, with their slow doubling rate
of 24 hours or more and relatively high cost of maintenance
due to more stringent sterility and growth requirements,
compounded by the concerns that most of them are
15 transformed cell lines, such mammalian cell lines have not
become the hosts of choice.

Insect cell lines, infected with recombinant
baculoviruses expressing antibody genes have also been
tested with some success, but despite having an efficient
20 signal sequence, about 50% of the total product has been
found to be retained within the cell. The use of insect
larvae, which have been demonstrated to be high producers
of recombinant intact antibodies, have been limited due to
concerns about potential contamination with bacterial
25 endotoxin beyond acceptable levels. The problems described
above have created a strong need for alternative methods
using, preferably, other eukaryotic host(s) for large-scale
production of intact antibodies and for consequent
reduction of traditional dependence on animals as sole
30 source of antibodies (Ibid, 1995).

It would, therefore, be advantageous to provide some
other biological system(s) capable of producing intact
antibodies which would be practical, economical, faster and
safer than these systems discussed above.

35 Yeast has a long history as a favorite host for

recombinant protein production, because of the unique advantages it offers as a unicellular eukaryote. Traditionally, the baker's yeast, *Saccharomyces cerevisiae*, was found to be suitable and is used as host for expression of recombinant proteins (Biotechnology, 9:1067 (1991)) including antibodies and Fab fragments PNAS USA, 85:8678 (1988)). However, despite some initial successes, it has not been possible to harness the full potential of *S. cerevisiae* for secreted production of intact antibodies (10 Nature Biotechnology, 16:773, (1998)).

In recent years, the methylotrophic yeast, *Pichia pastoris* (*P. pastoris*) has emerged as a popular host for over-production of both intracellular and extracellular recombinant proteins, including antibody fragments (J. Biochem., 121:831, (1997); and Bio/Technology, 13:255, (1995)).

Dioxins (halogenated dibenzodioxins) are highly persistent environmental contaminants with a broad spectrum of serious health effects, and there is a strong need for accurate detection of these toxicants (Nature, 375:353, (1995)). Since large quantities of antibodies are required for immunoassay in general and for rapid detection of dioxins in particular, the feasibility of using *P. pastoris* for producing functional, intact antibody against the prototypical dioxin, i.e., against 2,3,7,8-tetrachloro-dibenzo-p-dioxin (TCDD) was investigated and the current method for production of large quantities of hapten-specific antibodies secreted by *P. pastoris* was discovered. (25

It is, therefore, a subject of this invention to provide an antigen-specific intact recombinant antibody that is functional and a method for large scale production of such antibody. For that purpose, *P. pastoris* was evaluated as a host for efficient production of a recombinant monoclonal antibody (mAb), and large quantities of the intact recombinant antibody with binding-specificity (35

to dioxin, as a model, were produced.

The invention, therefore, concerns a large scale production and efficient secretion of a functionally assembled antigen-specific immunologically active intact recombina-
5 nt antibody with binding specificity to the antigen of interest by *P. pastoris*.

All patents, patent applications and publications cited herein are hereby incorporated by reference.

SUMMARY

10 One aspect of the current invention is a functionally assembled antigen-specific, immunologically active intact recombina-
nt antibody produced by transformation of the methylotropic yeast, *P. pastoris*, with human, mouse or other mammalian immunoglobulin genes.

15 Another aspect of the current invention is a method for production of functionally assembled antigen-specific intact recombina-
nt antibody by transformation of *P. pastoris* with human, mouse or other mammalian immunoglobulin genes.

20 Still another aspect of the current invention is the *P. pastoris* integrative expression vector (pPICZ α) into which particular antibody clones are subcloned in a two-
step process and the plasmid pPICZ α DD1 (for anti-dioxin antibody) or other pPICZ α recombina-
25 nt (depending on the antigen), as desired, is produced.

Still another aspect of the current invention is the transformation of *E. coli* XL1-Blue with the recombina-
nt plasmid of pPICZ α DD1 or with other pPICZ α recombinants.

BRIEF DESCRIPTION OF DRAWINGS

30 Figure 1 is a schematic representation of expression cassettes of the plasmid pPICZ α DD1. The expression cassettes of the 666-bp chain (L-chain) and 1332-bp heavy chain (H-chain) genes were each fused to the 267-bp yeast α -factor signal sequence (ss), under the control of the
35 yeast promoter (AOX1-P). A yeast transcription sequence

(TT) marks the 3' end of each expression cassette. The restricted enzyme sites used in the construction of the plasmid are indicated.

Figure 2 is a PCR analysis of *P. pastoris* transformants for pPICZ α DD1 genomic integrates. PCR products of *P. pastoris* transformants using primers specific for AOX1 and for the antibody light chain and heavy chain were analyzed using agarose electrophoresis.

Figure 3 is Northern blot analysis of RNA-blot of transcripts from clones 11501-1 (DD1) and 112535-1 (DD1) and two controls (ve+) and (ve-). The blots were probed with ³²P-labeled PCR amplicon of the antibody light chain and detected by autoradiography.

Figure 4 is Western blot analysis of culture media and cell lysates of recombinant yeasts probed with AP-goat anti-mouse IgG and visualized by AP color reaction.

Figure 5 is a graphic depiction of ELISA result demonstrating recombinant antibody binding and specificity to dioxin (hapten). The antibody-hapten binding was measured directly in an ELISA.

Figure 6 shows kinetics of antibody produced in yeast cells or secreted into cultured media, as revealed by immunoblots of cell lysates, and culture supernatants of *P. pastoris* probed with AP-goat anti-mouse IgG.

25 DEFINITIONS

As used herein:

"*Pichia pastoris*" or "*P. pastoris*" means a methylotropic yeast, a single-celled microorganism that prefers aerobic growth and can be grown to much higher cell densities than fermentative yeasts.

"DD1" means mouse hybridoma secreting antidioxin monoclonal antibody described in the U.S. patent 5,334,528.

"Antibody genes" means and is used to denote the mRNA, cDNA, or genomic or chemically synthesized DNA fragments coding for an antibody.

DETAILED DESCRIPTION OF THE INVENTION

This invention concerns and describes a novel method for production of functionally assembled antigen-specific, immunologically active intact recombinant antibodies. The method is generally useful for preparation of any kind of antibody and is also suitable for large-scale production of human and other mammalian antibodies. The method is easy, practical, fast and safe.

I. Method for Production of Functionally Assembled Antigen-Specific Intact Antibody

The method of the invention for production of functionally assembled antigen-specific intact monoclonal antibody, using transformation of *P. pastoris*, has a general utility and essentially any antibody can be produced or secreted by *P. pastoris* as long as the yeast expression vector carrying antibody genes can be appropriately assembled.

Briefly, *P. pastoris* is transformed with human, mouse or other mammalian immunoglobulin genes encoding heavy (gamma) and light (kappa or lambda) chains of antigen-specific antibody. Antibody genes are isolated from a hybridoma that recognizes certain specific antigen and the transformed yeast clones generated according to the method of invention then specifically recognize that particular antigen. Following the transformation, *P. pastoris* produces and secretes large amounts of a functionally assembled antigen-specific intact monoclonal antibody into the culture supernatant.

In a more detailed description of the method, a recombinant yeast expression vector (pPICZ α) with dual expression cassettes is constructed, each cassette carrying the inducible alcohol oxidase (AOX1) promoter, fused to the *Saccharomyces cerevisiae* α -factor signal sequence. *P. pastoris* is then transformed with these constructs, and the resulting transformant secretes functionally assembled

intact recombinant antibody molecules into the medium from where it is readily recovered using affinity purification procedures.

Specificity of the produced antibody is determined by demonstrating the antibody-specific mRNA synthesis in recombinant yeast using Northern blot analysis. When the specific antibody is produced, immunoblot and ELISA analyses of concentrated culture supernatants harvested a few days post-transformation reveal the presence of antigen-specific human, mouse or other mammalian species-specific immunoglobulins. Assaying of the culture supernatants by ELISA then shows specific binding activity to the specific antigen against which the antibody is raised or to a cross-reactive congener. The binding affinity of the produced recombinant IgG is the same as, and/or comparable to, that of the parent IgG.

II. General Method for Production of Antigen-Specific Intact Antibodies on Large Scale

The method according to the invention utilizing *P. pastoris*-based expression system was found suitable for a large-scale production of intact recombinant antibodies.

A typical process for large scale production of antigen-specific antibodies according to the invention comprises following steps:

Step 1. Selection and identification of the antigen against which the antibody is to be raised.

Such antigen selection is entirely need-based. If the aim is, for example, to produce an antibody against the AIDS virus, HIV-1, then inactivated HIV-1 is used as the immunogen. If the genes (or their cDNA) are available that code for anti-HIV-1 antibody, the method proceeds directly to step 2 of the procedure and those genes are expressed using the vector system and method or an appropriate variation thereof of the invention. If the aim is, for example, to produce antibodies that recognize the malaria

antigen (Ag), then the anti-malaria Ag antibody genes, if available, are used or these genes are isolated from a hybridoma secreting monoclonal antibodies that recognize and bind malaria antigen.

5 In yet another example of how to obtain the antibody, an animal is immunized with the target immunogen or carrier-conjugated hapten, a cDNA library of either the IgG repertoire or the entire mRNA component is generated and the library is screened for clones with specific
10 recognition of the target antigen. Similarly, the antibody genes may be isolated directly from a previously exposed or a naive animal or human and expressed using the expression system of the invention.

In the case of an immunogen/hapten that is a chemical
15 compound, it is either purchased or custom-ordered from a chemical company (e.g., Aldrich, Chemicon) or, if it is not available commercially, it is synthesized in laboratory. Likewise, protein or peptide immunogens/antigens can either be purchased from any of a number of biochemical companies
20 (e.g., Sigma Calbiochem, etc.), purified from target source, such as animal tissues/cells, plants, bacteria, viruses, etc., prepared from an existing precursor, such as peptides from a pre pro- or pro- or mature protein, or obtained by recombinant methods. Haptens and peptides that
25 are too small to be effective as immunogens are used in conjunction with suitable carriers for production of antibody, generally monoclonal antibody, and the antibody-encoding genes are isolated from the antibody-producing cells and used in the *Pichia* expression system for creating
30 a source of candidate antibody as well as antibody genes.

Step 2. Isolation or chemical synthesis or PCR-/T7-amplification of antibody-encoding cDNA, mRNA or genomic DNA fragments.

Methods for isolation and chemical synthesis or PCR or
35 T7 amplification of cDNA, mRNA or genomic DNA are known in

the art. Any and all these methods may be used for the step 2 of this method.

Step 3. Assembling the antibody genes into expression cassettes.

5 Assembling the antibody genes into expression cassettes, as seen in Figure 1 is achieved by, for example, subcloning the light and heavy chain cDNAs in tandem as *EcoRI-BglIII/BsmBI* fragments each flanked by a signal sequence, such as, for example yeast α -factor, preceded by
10 a yeast promoter, such as for example alcohol oxidase AOX1-P, at the 5'-terminus and by the yeast transcription termination sequence at the 3'-terminus. Possible variations of this step include use of antibody genes such as the use of the entire light chain (kappa or lambda),
15 partial use of light chain comprising only one, two or three CDRs or parts thereof, one or two CDRs in combination with part or complete framework region (homologous, heterologous or non-Ig but neutral compatible sequence from a diverse source such as myoglobin, actin or a synthetic
20 peptide of unrelated origin and/or function) and/or a heavy chain gene of similar variations. Other variations include use of signal and promoter sequences, including but not limited to those obtained from insects, yeasts, bacteria, viruses, mammals, and plants, as long as they are
25 functional in *P. pastoris*.

Step 4. Preparing a yeast expression vector pPICZ α for cloning of antibody genes.

A yeast expression vector, for example, pPICZ α is prepared for cloning of antibody genes, for example, by
30 restriction digestion with *EcoRI* and *BamHI*. Other restriction enzymes unique to the vector or such that if they were present in the antibody genes they can be repaired to restore the functional integrity of the antibody genes in the recombinant plasmid may be also used
35 for preparation of the vector.

Step 5. Cloning of antibody gene expression cassettes into the *Pichia* expression vector (pPICZ α).

Antibody gene expression cassettes are cloned into the *Pichia* expression vector (pPICZ α) to generate recombinant plasmid (pPICZ α LH) or a variant thereof using methods known in the art for cloning.

Step 6. Transforming bacteria with recombinant plasmid pPICZ α LH or its variant.

Bacteria, for example *Saccharomyces cerevisiae* are transformed with recombinant plasmid pPICZ α LH or its variant using methods known in the art.

Step 7. Amplifying and isolating the recombinant plasmid on preparative scale.

Amplification and isolation of the recombinant plasmid on preparative scale is achieved by using standard methods of large-scale growth of the recombinant and plasmid isolation such as the alkaline lysis method described in Current Protocols in Molecular Biology, Ausubel E.M. et al., Wiley-Interscience, New York, (1990).

Step 8. Preparing and transforming *P. pastoris* spheroplasts.

P. pastoris spheroplasts are transformed with *Bgl*III-linearized, or in alternative *Not*I, *Sac*I, *Sal*I and *Stu*I-linearized recombinant plasmid, resulting in *in vivo* homologous recombination replacement of the yeast chromosomal AOX1 sequence with the 5' AOX1-antibody gene cassette of the recombinant plasmid.

Step 9. Selectively growing the recombinants and eliminating the non-recombinants.

Selective growth of recombinants and elimination of non-recombinants is achieved by plating transformants on medium containing zeocin (100 ug/ml). Zeocin may be replaced by other compounds. Such replacement depends on the selection marker (gene) included in the plasmid. Some examples of possible replacement include, but are not

limited to, G418, trimethoprim, and drugs/compounds/polypeptides that limit the growth of wild type *P. pastoris* in contrast to the yeast that is transformed with a plasmid containing said selection gene(s).

Step 10. Screening of the yeast transformant colonies for antibody expression.

Screening of the yeast transformant colonies for antibody expression is achieved by colony-immunoblotting for the origin of the recombinant proteins (antibody): human/mouse/or other; antigen-/hapten-binding activity thereby providing preliminary identification of putative positive clones. In alternative, any other means of distinguishing the recombinant over the host background may be used.

Step 11. Analyzing the putative positive yeast clones for chromosomal integrates of the expression cassettes.

The putative positive yeast clones are analyzed, for example, by PCR or by restriction analysis, for chromosomal integrates of the expression cassettes of both light and heavy chain cDNAs at the correct locus.

Step 12. Performing a Mut⁺/Mut^s test for selecting +His + Mut⁺ phenotypes.

Performing a Mut⁺/Mut^s test by replica-plating transformant colonies on (i) a -His +glucose plate and on (ii) a -His +methanol plate and, because Mut⁺ colonies are slow utilizers of methanol while Mut⁺ colonies are normal, that is relatively rapid, utilizers of methanol facilitated by AOX1 promoter, selecting +His + Mut⁺ phenotypes.

Step 13. Confirming the DNA insert/junction sequence integrity.

The DNA insert/junction sequence integrity is confirmed by nucleotide sequence analysis using standard methods of DNA sequencing, such as the chemical sequencing method or the dideoxy termination method including the

automated methods (Current Protocols in Molecular Biology, Ausubel F.M. et al., Wiley-Interscience, New York, (1990)).

Step 14. Inducing recombinant antibody expression and growth.

5 Inducing recombinant antibody expression and growth by, for example, methanol [0.5 to 1.5%, v/v] and glycerol [1%, v/v]) at 30°C or any other conditions including potential gratuitous inducers and other growth conditions that elicit induction of antibody.

10 Step 15. Establishing the antibody authenticity.

The antibody authenticity is established, for example, by Northern blot/RNA protection analysis of the clones.

Step 16. Detecting the presence of the recombinant antibody.

15 The presence of the recombinant antibody is detected, for example, by Western blot analysis of the yeast cellular proteins and proteins secreted into the culture supernatant of the yeast clones.

Step 17. Demonstrating the antibody-antigen-specific
20 binding activity.

Specificity of recombinant antibody specific antigen-antibody binding is confirmed, for example, by ELISA or other methods that recognize antigen-antibody reaction.

Step 18. Optimizing recombinant antibody production.

25 Optionally, the production of recombinant antibody is optimized by testing a broad range of culture and induction conditions.

Step 19. Purifying and storing the recombinant antibody.

30 Optionally, the recombinant antibody is purified and stored under conditions that favor its optimal stability and recovery, by for example, storing the antibody in the presence of protease-inhibitors, at -80°C or in the presence of cryoprotective agents such as 50% glycerol.

35 The method described above allows expression of cDNA

fragments encoding antibody light and heavy chains isolated from a pre-existing hybridoma, as illustrated below in Section III, for the preparation of the anti-dioxin antibody. Anti-dioxin antibody and hybridomas for their expression are described in US patent 5,334,528 hereby incorporated by reference for method of producing hybridomas. The anti-dioxin genes were isolated from the hybridomas DD1 or DD3 and were genetically engineered into the *Pichia* expression system and coordinately expressed, producing immunologically active intact recombinant antibody.

The process for the preparation of any antigen-specific antibody according to the invention is able to utilize any existing hybridoma. Once the light and heavy chain of the antibody cDNAs are isolated by one of the standard methods known in the art, the ends of the cDNA fragments are modified to match any one or more of the multiple cloning sites (*Bam*HI, *Sna*BI, *Eco*RI, *Avr*II and *Not*I), and cloned into that site in a vector of the pPICZ family.

The same approach is useful for genomic PCR amplicons or clones coding for antibody light chain and heavy chain open reading frames (ORFs).

The process is further useful and applicable to any available mammalian hybridomas, as well as to human hybridomas, as long as appropriate primer sequences are designed or selected for sequence amplification. Moreover, once the antibody genes are cloned into a *Pichia* expression vector, the rest of the scheme (Steps 6 through 19) is the same as described above but for the antigen-specific reagents.

The method is generally useful also for expression of antibody genes isolated from clinically and industrially important hybridomas producing monoclonal antibodies to c-myc, Her2/neu, lymphoma, etc., or for cloning and

expression of antibody genes from immunized or naive animals or humans. The only variations in the method apply to cloning of the antibody genes. Resulting clones determine the properties of the antibody that is eventually
5 produced using this approach.

If PCR amplification, for example, is used for cloning the antibody genes, then the primer sequences used for PCR amplification determine the antibody type, form and size. In other words, the primers can be designed to isolate
10 genes from an animal or from a human, and to produce an intact antibody or a fragment such as Fab.

If gene probes are used for isolating antibody genes, then the antibody that is produced from them is dependent on the probes used. If probes are specific for a human
15 antibody, then the recombinant antibody that is expressed from those genes is the human antibody. On the other hand, if the probes are specific to a mouse antibody then the resulting recombinant antibody is the mouse antibody. However, by capitalizing on the relatively close homology
20 between the mouse and human gene sequences which permits generation of so called humanized mouse, it is possible to use gene probes to isolate antibody gene clones for cross-species by simply reducing the stringency of the probe hybridization conditions.

Further, if the sequences of the primers or probes are specific, for instance against mouse c-myc antibody by recognizing unique sequences in their CDRs (complementarity determining regions) and not the framework regions (common to different antibodies), then the cloned antibody genes
25 will code only for c-myc antibody and not any other antibody. By the same token, when the primers or probes are chosen to recognize the framework regions of the antibody chains, a heterogeneous population of antibody genes is obtained.
30

35 One precondition for the method is a selection of a

single pair of heavy and light chain genes either at the time of constructing recombinant yeast expression plasmid or at the time of isolating individual yeast clones expressing recombinant antibodies. Typically, to clone
5 antibody genes directly from an animal or a human, the genes for any antibody can be isolated and expressed. There is no other limitation within the method, except for the necessity of designing the primers or probes or screening the libraries of clones by using methods known in the art.

10 The procedure for producing any antibody is thus essentially the same as the one described above for general purposes, and below, as exemplarized for recombinant anti-dioxin antibody, said procedure needing only to suitably incorporate the variations described for steps 1-5.

15 III. Method for Production of Functionally Assembled Anti-Dioxin Specific Intact Antibody

The method of the invention was developed and tested on anti-dioxin antibody because the anti-dioxin antibody (IgG gamma and kappa) genes are readily available in the
20 inventors laboratory and also because, as a practical matter, a large quantities of the anti-dioxin antibody was urgently needed for fast and reliable detection of dioxin contamination in the environment.

In the following description of the procedure used for
25 production of anti-dioxin antibody according to the invention, subsections A-E deal with the specific description of the method for production of anti-dioxin antibody, including steps, procedures, materials and test results.

30 A. Anti-Dioxin Antibody Synthesized by *P. pastoris* Transformed with pPICZ α DD1

The invention is based on the discovery that the methylotropic yeast *P. pastoris* can be transformed to secrete large quantities of a dioxin-specific antibody when
35 transformed with *Pichia* expression vector carrying the

genetic information for expression of anti-dioxin antibody.

1. Methylootropic Yeast *Pichia pastoris*

P. pastoris strain SMD1168 (pep4 his4) was identified as a suitable host for antibody production according to the invention.

P. pastoris strain SMD1168 (pep4 his4) and the *P. pastoris* integrative expression vector (pPICZ α B) were obtained from Invitrogen (Carlsbad, CA).

2. Transformation of *Pichia pastoris*

The *P. pastoris* strain SMD1168 was transformed with recombinant plasmid, pPICZ α DD1 and zeocin-resistant transformants were isolated.

A schematic representation of expression cassettes of pPICZ α DD1 plasmid is shown in Figure 1. The 7164-bp recombinant plasmid, pPICZ α DD1, contains a bacterial origin of replication (COIE1), Zeocin-resistance gene (Zeo^R) for selection of both *E. coli* and yeast transformants, and the expression cassettes of anti-dioxin antibody light- and heavy chain genes.

The final construct was assembled by replacement-ligation of the 3134-bp (*Mlu*I-*Bam*HI) fragment of the recombinant plasmid construct containing the light chain expression cassette, and the 4030-bp (*Mlu*I-*Bam*HI) fragment from the heavy chain expression cassette construct.

Figure 1 is a schematic representation of expression cassettes of the plasmid pPICZ α DD1. The expression cassettes of the 666-bp light-chain (L-chain) and 1332-bp heavy-chain (H-chain) genes were each fused to the 267-bp yeast α -factor signal sequence (SS), under the control of the yeast promoter (AOX1-P). A yeast transcription termination sequence (TT) marks the 3' end of each expression cassette. The restriction enzyme sites used in the construction of the plasmid are indicated.

DNA manipulations were performed using standard techniques as described in Current Protocols in Molecular

Biology, Eds. Ausubel, F.M., et al., Wiley-Interscience, New York, (1994) or according to vendor recommendations.

The light-chain (666-bp) and heavy-chain (1332-bp) sequences of the mouse hybridoma DD1 (Gene, 19:388, (1994))
5 secreting anti-dioxin mAb, have been cloned and sequenced and expressed in *E.coli* as Fab according to J. Agric. Food Chem., 46:3381 (1998).

The anti-dioxin antibody genes were sub-cloned into pPICZ α B in a two-step process. First, the sequence
10 encoding the light-chain (666 bp) or heavy-chain (1332 bp) mature peptide was PCR-amplified from cloned cDNA using primers designed to produce a blunt 5' terminus and a 3' nested *Bgl*III/*Xba*I site preceded by a stop codon, and a codon for cysteine (TGC) introduced before the stop codon
15 in the heavy-chain sequence to facilitate conjugation of the recombinant antibody to a peptide tag for affinity purification.

The light-chain and heavy-chain amplicons were separately cloned into pPICZ α B, under the control of yeast
20 AOX1 promoter, translationally fused to *S. cerevisiae* α -factor signal sequence, producing pPICZ α -L (light) and pPICZ α -H (heavy) plasmid, respectively.

In the second step, plasmid pPICZ-L was digested with *Bam*HI + *Mlu*I and pPICZ α -H with *Bgl*III + *Mlu*I, and the 3134-
25 bp vector fragment containing the light-chain expression cassette and the 4030-bp vector fragment containing the heavy-chain expression cassette, were gel-eluted. The final 7164-bp construct pPICZ α DD1 was assembled by replacement-ligation of the gel-eluted light-chain and
30 heavy-chain fragments.

Detailed method and transformation conditions for introduction of the pPICZ α DD1 construct into *E. coli* are described in Example 2.

3. Screening For Antibody Expression

35 Screening of transformant yeast colonies for antibody

expression was performed as described in Example 3.

Briefly, the zeocin-resistant yeast colonies were patched on nitrocellulose filters and grown for 2 days on induction plates at 30°C. The colony-blot was probed with
5 AP-goat anti-mouse monoclonal antibody (Boehringer Mannheim, Indianapolis, IN) as recommended by the vendor.

DNA sequencing procedure is described in Example 4. DNA was processed for sequencing using the ABI Tag DyeDeoxy Terminator Cycle Sequencing kit, based on the chain
10 terminating method. Nucleotide sequence was determined in an automated DNA Sequencer and data analyzed using the PE/ABI editing and assembly software.

Transformants for antibody expression were screened by PCR analysis as described in Example 5. Results are shown
15 in Figure 2.

Screening of the transformants for antibody expression was performed by preparing a nitrocellulose membrane-replica of transformants growing on an agar plate with induction medium and probing it with anti-mouse IgG (data
20 not shown).

Two transformants, 11505-1 (Mut⁻) and 112535-1 (Mut⁺) that tested strongly positive by this screen were analyzed by colony-PCR using relevant primers to confirm chromosomal integration of the plasmid DNA sequences.

Figure 2 shows PCR analysis of *P. pastoris* transformants for pPICZαDD1 genomic integrates. PCR products of *P. pastoris* transformants using primers specific for AOX1 are shown in lanes 2-7, the antibody light chain are shown in lanes 8-11 and heavy chain in
30 lanes 12-15. Transformants were analyzed using agarose (1.5%, w/v) gel electrophoresis. Figure 2, lane 1 shows Mr. markers (Phage lambda BstEII digest; New England BioLabs, Beverly, MA); lanes 2, 8, and 12 show clone 11505-1; lanes 3, 9, and 13 show clone 112535-1; lanes 4, 10, and
35 14 show unmodified vector (-ve control); lanes 5-7, 11, and

15 show recombinant plasmid (+ve control); lane 5 shows light-chain gene; lane 6 shows heavy-chain gene; lanes 7, 11 and 15 show vector with both light- and heavy-chain genes; lane 16 shows Mr. markers (PhiX174 HaeII digest; New England BioLabs). Molecular weights of the markers are shown in kilobases (kb) on the left of the gel and in base pairs (bp) on the right.

Results obtained with these two representative clones, shown in Figure 2, confirmed the integrity of expression cassettes in the 11505-1 and 112535-1 clones by DNA sequence analysis.

PCR analysis of *P. pastoris* transformants using primers specific for the antibody genes or for the AOX1 5' and 3' termini showed intact full-length light- and heavy-chain gene expression cassettes integrated in genomic DNA of the transformants. Control transformants harboring vector alone yielded no amplification products in PCR with Ab gene primers. DNA sequence analysis of the PCR products from recombinants confirmed that the primary structure of the target sequences was preserved.

4. Induction of Antibody Production

Following the confirmation and integrity of expression cassettes of the clones, the antibody production by recombinant *P. pastoris* clone was induced according to Example 6.

Induction of recombinant antibody expression was typically performed as follows. A *P. pastoris* transformant was cultured for two days with shaking at 250 rpm in BMGY broth (buffered glycerol-complex medium with yeast extract) at 30°C. The yeast cells were collected by centrifugation and transferred to the induction medium. Beginning on the second day of growth, methanol was added daily to a concentration of 0.5% (v/v), to induce the AOX1 promoter-driven production of recombinant antibody.

Screening of the recombinants for antibody expression

was performed using the colony-blot assay (data not shown). The method involved making a nitrocellulose membrane-replica of recombinants on an agar plate with induction medium and probing it with AP-goat anti-mouse IgG, after
5 gently washing the cell-debris off with non-fat milk (5%, w/v, in TBST).

Fifteen recombinants showing a high degree of reactivity, indicating potential for high levels of recombinant Ab production, were picked for further
10 analysis. From among those that tested positive by colony-PCR, diagnostic restriction digestion and DNA sequence analyses identified two clones as clones 11505-2 and 112535-1. These two clones were chosen for further characterization.

15 Specificity of transcripts was determined by Northern blot analysis according to Example 7.

For Northern blotting, total RNA was extracted from both clones (11505-1 and 112535-1) and the vector control, each induced in a 5-ml culture in MMH medium at 30°C for 4
20 days.

RNA (20 μg sample⁻¹) was denatured, resolved by 1% agarose gel electrophoresis in 1 x MOPS buffer containing formaldehyde 1.2%, transferred to a nylon membrane (Hybond-N; Amersham Pharmacia Biotech, Piscataway, NJ) and
25 probed with ³²P-labelled light-chain amplicon. The blot was washed, air-dried and exposed to a Kodak X-O-matic film for 24 hour at -80°C. Results are shown in Figure 3.

Figure 3 is Northern blot analysis of total RNA transcripts from the two (11505-1 and 112535-1) clones and
30 two controls (one positive and one negative) probed with ³²P-labelled PCR amplicon of the antibody light chain and detected by autoradiography. Lane 1 shows a light-chain amplicon (+ve control), lane 2 shows clone 11505-1; lane 3 shows clone 112535-1; lane 4 is a vector (-ve control).
35 The RNA Ladder (New England BioLabs, Beverly, MA) stained

with ethidium bromide was used as reference for estimating RNA sizes.

Transcripts detected in the Northern blot as seen in Figure 3 were specific and corresponded to the sizes expected for both light- and heavy chains. Although the probe was derived from the light chain, it also recognized the heavy-chain transcript, because of the partial homology shared between the two chains. The variation observed in the strength of signals produced in Northern blot by different samples reflected the difference in relative levels of antibody expressed by the clones analyzed. By this measure, clone 11505-1 (lane 2) expressing the DD1 antibody showed greater expression levels than 112535-1 (lane 3).

As seen in Figure 3, Northern blots of total RNA from induced (96 hours) cultures of the clones 11505-1 (lane 2) and 112535-1 (lane 3), probed with gel-purified and labeled light-chain gene, showed specific transcripts of 1360 bp and 2022, corresponding to the sizes expected for the light- and heavy-chain genes, respectively.

B. Efficiency of Intact Antibody Secretion by *Pichia Pastoria*

The intactness of the inserts and the accuracy of the junction sequences were confirmed using PCR procedure described in Example 9 and by nucleotide sequence analysis. For Western immunoblot analysis, cell lysates and 25 x concentrated media containing between 50-75 μ g a total protein sample⁻¹, were resolved by non-reducing 10% SDS-PAGE using Tris-glycine SDS buffer and electroblotted onto a PVDF membrane (Millipore, Bedford, MA). The blot was processed and probed with AP-goat anti-mouse IgG (1:5000, in TBST, pH 8.0) using BCIP and NBT. Results are shown in Figure 4.

Figure 4 shows Western blot analysis of culture media and cell lysates of recombinant yeasts. Nitrocellulose

blots containing equivalent amounts of total yeast proteins from culture media and cell lysates from *P. pastoris* clones and controls were probed using AP-goat anti-mouse IgG and were visualized by AP color reaction. For Figure 4, H₂L₂ shows intact Ab, HL is a heavy chain-light chain monomer; H is gamma (heavy) chain; L is kappa (light) chain. Lane 1 shows prestained protein Mr markers (New England BioLabs) with sizes indicated on the left of the gel, lane 2 shows clone 11505-1; lane 3 shows clone 112535-1, lane 4 shows vector (-ve) control and lane 5 shows mouse IgG, 0.25 µg (+ve control).

Western-blot analysis of culture media of the clones 11505-1 and 112535-1 revealed mouse antibody chains with the sizes expected for monomers of light chain (25 kDa), heavy chain (50 kDa), intact antibody (150 kDa) and some intermediate assemblages as seen in Figure 4, indicating proper assembly of antibody molecules. Results of non-reducing SDS-PAGE analysis of culture media and cell lysates of the clones 11505-1 and 112535-1 (data not shown) corroborated immunoblotting results seen in Figure 4.

Antibody levels in cell lysates of both clones (11505-1 and 112535-1), at approximately 10% of the total product, as assessed by Western blotting, were consistently lower than those found in culture medium, demonstrating that a major portion, approximately 90% of the antibody produced by recombinant *P. pastoris* was secreted into the supernatant. These results show that *P. pastoris* secretes intact antibody very efficiently. Non-reducing SDS-polyacrylamide gel electrophoresis (SDS-PAGE) of culture media and cell lysates showed protein bands migrating at positions expected for the light (25 kDa) and heavy chain (160 kDa) assembled antibody.

C. Specificity of the Recombinant Antibody

The binding specificity of the recombinant antibody to

its cognate hapten, in this case dioxin, was evaluated using two independent approaches, slot-blot and ELISA described in Examples 9 and 10.

5 The hapten-binding activity of recombinant antibody was assayed using ELISA. Briefly, 96-well microtiter plates (MaxiSorp Nunc-Immuno Plate, Nalge Nunc International, Denmark) coated with 10-50 ng range of BSA-dioxin, trans-3-(2,3,7, 8-tetrachlorodibenzo-p-dioxin-1-yl) propenoic acid, in 100 μ l well⁻¹ of 50 mM bicarbonate
10 buffer, pH 9.6, were incubated with 100 μ l of a 1:10 dilution (in TBST) of culture medium from an induced culture of each recombinant and were probed with HRP-goat anti-mouse IgG (Pierce Chemical, Rockford, IL) using the chromogenic TMB Microwell Peroxidase Substrate System (KPL, Gaithersburg, MD). The plates were read at 450 nm, using
15 the UVmax kinetic microplate reader (Molecular Devices, Menlo Park, CA), and the readings were analyzed using the software package Softmax (Molecular Devices, Menlo Park, CA).

20 The hapten-binding activity of the recombinant antibody was analyzed using slot-immunoblotting. All incubations were performed at 26-28°C. The hapten (BSA-dioxin, diluted 1:3600) was spotted in 10 μ l volumes (to give 250 ng slot⁻¹) on a nitrocellulose membrane (S&S, Keene, NH), using a slot-blot apparatus (Life Technologies,
25 Rockville, MD). The blot was allowed to air-dry, blocked with non-fat milk (10%, w/v, in TBST) for one hour, and incubated with gentle shaking (120 rpm) on an orbital shaker (Lab-Line, Melrose Park, IL) for one hour with 2 ml
30 of 1:10 dilution in TBST of the medium from induced cultures of the clones. The blot was washed, incubated with AP-goat anti-mouse IgG and developed as described for the Western immunoblot.

Both approaches demonstrated specific binding of the
35 recombinant antibody to cognate hapten, dioxin, with

affinities comparable to those of the parental mAb, DD1. These values can be affirmed only after precise quantification, of protein concentrations of the recombinant antibody after accounting for the *Pichia* cellular proteins secreted into medium. Results are seen in Figure 5.

Figure 5 is a graphic depiction of ELISA result demonstrating recombinant antibody binding to dioxin. The antibody-hapten binding was measured directly by ELISA. The coating hapten, BSA-dioxin (10 to 40 ng well⁻¹) in a 96-well microtiter plate was incubated with serial dilutions of culture medium from clones 11505-1 and 112535-1, vector (-ve) control, or DD1 mAb (+ve control), and probed with HRP-goat anti-mouse IgG using TMB. The A₄₅₀ readings indicate the hapten-binding activity of the recombinant antibody. Results shown in Figure 5 demonstrate that the produced recombinant antibody is functional.

D. Kinetics of Monoclonal Antibody Production

Studies performed to determine the kinetics of monoclonal antibody have shown that synthesis and secretion of antibody are optimal between 72 and 108 hours. Detailed procedure is described in Example 10. Results are shown in Figure 6.

Figure 6 illustrates kinetics of antibody production or secretion. Slot-blots of immobilized cell lysates or culture media from the clone cultures harvested in 12-hour intervals of induction (12 to 120 hours) were probed with AP-goat anti-mouse IgG. The clones and the duration of induction, in hours, are indicated. Figure 6A shows culture media (supernatant), where top row shows clone 11505-1 and bottom row shows clone 112535-1. Figure 6B shows cell lysates, where top row shows clone 11505-1 and bottom row shows clone 112535-1. Figure 6C top row shows vector (-ve) control and bottom row shows mouse IgG (+ve control).

The kinetics of antibody production/secretion were followed by withdrawing portions of the culture at various intervals of methanol-induction of the clones and determining the antibody levels by slot-immunoblot analysis. As seen in Figure 6, anti-dioxin antibody was detectable in culture medium between 12 hours and 120 hours of induction, with highest levels of about 10 to 36 mg l⁻¹ detected between 72 and 108 hours.

Although these levels are lower than those reported for other recombinant proteins or for antibody fragments (200 mg l⁻¹), they are the highest and set the highest range ever obtained for any intact antibody, which is a more complex multimeric glycoprotein, than the molecules previously reported. These levels can be further augmented by using fermentation approaches.

UTILITY

This invention provides a method of general utility for production of large quantities of any antigen-specific antibody using modified yeast organism. Using the method of invention, the large quantity of compound specific and defined monoclonal antibody is produced without the necessity of immunizing and recovering and purifying antibodies and/or other lengthy procedures. The method is practical, economical, easy, safe and fast and in about three days, the monoclonal antibody is produced by the transformed yeast if the vector and expression vehicles for transformation are available or are prepared according to the invention.

The above described findings demonstrate the suitability of *P. pastoris* expression system for both small and large-scale production of functional, antigen-specific intact antibodies. The recombinant antibodies produced by the method of the invention are useful, for example, for immunodiagnostic and immunotherapeutic purposes. Since recombinant proteins produced in *P. pastoris* lack terminal

α 1,3 glycan linkages responsible for hyper-immunogenicity, the antibodies produced in *P. pastoris* are particularly suitable for therapeutic applications.

Functional assembly of antibodies produced in *P. pastoris* also suggests the potential of *P. pastoris* for construction of antibody libraries and screening them with any antigen of interest using colony-immunoblotting.

EXAMPLES

Materials

10 Restriction enzymes were purchased from New England BioLabs (Beverly, MA), and Taq Polymerase from Promega (Madison, WI).

cDNA clones of the heavy and light chains of the anti-dioxin mouse mAbs, DD1 and DD3, were a gift from A. Recinos
15 III and L. Stanker. Primers, designed on the basis of the nucleotide sequence of the above cDNAs, were synthesized by Life Technologies (Gaithersburg, MD).

96-well microtiter plates (Nunc-Immunoplate, Maxisorp) were from Nalge Nunc International (Roskilde, Denmark), and
20 HRP-goat anti-mouse IgG was obtained from Pierce Chemical (Rockford, IL). All chemicals were of reagent grade from Fisher Scientific (Pittsburgh, PA) or from Sigma (St. Louis, MO).

EXAMPLE 1

25 Microbial Strains and Culture Conditions

This example identifies microbial strains and culture conditions used for the purposes of this invention.

Escherichia coli strain XL 1-Blue was used as host for plasmid amplification, using YB broth (1.5% tryptone, 1%
30 yeast extract, 0.5% NaCl). *P. pastoris* SMD1168 (pep4 his4) and the yeast expression vector (pPICZ α B) were obtained from Invitrogen (Carlsbad, CA).

The yeast was grown in minimal dextrose medium obtained from DIFCO (Detroit, MI), supplemented with
35 histidine (MDH:1.34 % YNB without amino acids, 4×10^{-5} %

biotin, 2 % dextrose, 0.004 % L-histidine) and was induced in MMH medium (minimal methanol medium supplemented with histidine: 1.34 % YNB, $4 \times 15 \times 10^{-5}$ % biotin, 1.5% methanol, 0.004% L-histidine).

5

EXAMPLE 2Construction of Expression Plasmid

This example describes construction of the expression plasmid.

Complementary DNAs (666 bp and 1332 bp of light and heavy chains, respectively) anti-dioxin genes were cloned separately into a PPICZ/ α *P. pastoris* integrative vector with zeocin resistance gene. For the cloning, the genes were placed under the control of AOX1 promoter alongside of α -factor signal sequence using the *EcoRI* ends blunt-ended with T4 polymerase prior to digesting with *BsmBI* using methods known in the art.

The PCR primers were synthesized with a *BglIII* site incorporated at the end of the stop codon, the product of the cDNA was cloned through *BglIII* site ligated into a *BsmBI* site of the vector, resulting in the loss of both sites in the recombinant plasmid generated.

The individual recombinant plasmids were then digested with *BamHI* and *MluI*, for the recombinant containing the light chain and with *BglIII* and *MluI* with the heavy chain construct. From each construct a 3134-bp fragment representing the light chain and 4030-bp fragment representing the heavy chain constructs were gel-eluted and religated to contain both the light and heavy chain genes.

The construct was introduced into *E. coli* XL1-Blue by electroporation, and recombinants were selected by scoring for zeocin (25 mg/ml) resistance. DNA was extracted and purified from recombinants, confirmed by colony PCR, linearized with *DraI* and used for transforming *P. pastoris* SMD1168 by electroporation using Gene Pulser (Bio-Rad, Richmond, CA).

Cells were regenerated in ice-cold 1 M sorbitol (1 ml) at 30°C for 2 hours, plated in 10-100 μ l portions on YEPD medium (1% yeast extract 2% peptone, 2% dextrose) containing zeocin (100 μ g/ml) and incubated at 30°C for 3-5 days. DNA manipulations were all performed using standard techniques or as recommended by the respective reagent vendors.

Recombinant colonies were each screened for the presence of genome-integrated inserts using colony-PCR, and for growth, at 30°C for 3-4 days on Minimal Methanol (MM) medium.

EXAMPLE 3

Expression-Screening of Transformants

This example describes procedure used for screening of transformants.

The yeast colonies, which grew on zeocin selection, were replica-plated on MM agar plates and incubated for 2 days at 30°C, colonies were covered with nitrocellulose membrane and allowed to grow further for 2-3 days at 30°C. The membranes with yeast colonies were washed 3 x with TBST, blocked for 1 hour with nonfat dry milk (10% w/v) in TBST, and incubated with Alkaline Phosphatase-conjugate (AP-goat) of goat anti-mouse monoclonal antibody (Boehringer Mannheim, IN, USA) diluted 1:5000 in TBST. After 1 hour, the membranes were washed 5 x with TBST and developed in the dark for 10-30 minutes at room temperature in 100 mM Tris-HCl, pH 7.5, 50 mM NaCl and $MgCl_2$ containing the chromogenic substrates, NBT and BCIP.

EXAMPLE 4

DNA Sequencing

This example describes DNA sequencing protocol.

The nucleotide sequence of the constructs was determined using the chain terminating method according to PNAS (USA), 74:5463 (1977) of DNA sequencing using the ABI Taq DyeDeoxy terminator cycle sequencing kit on an

automated DNA Sequencer (ABI 373A, Applied Biosystems, Foster City, CA). The sequences were analyzed using the ABI Prism software for sequence analysis.

EXAMPLE 5

5 PCR Analysis of Expression Cassettes

This example describes PCR analysis of expression cassettes.

Genomic DNA was isolated from transformed and control (non-transformed) yeast cells, and 400 ng was tested for
10 the presence of expression cassettes by PCR analysis using the following specific oligonucleotide primers for

the light-chain gene:

Forward: 5'-GACGTCGTGATGACCCAAGCTCCA-3' (SEQ ID NO:1)

Reverse: 5'-CGCGTCTAGATCTAACACTCATTCTGTTGAA-3' (SEQ ID NO:2)

15 the heavy-chain gene:

Forward: 5'-CAGGTCCAAGTGCAGCAG-3' (SEQ ID NO: 3)

Reverse: 5'-CGCGTCTAGATCTAGCATTTACCAGGAGAG-3' (SEQ ID NO: 4)

the yeast AOX1 promoter

Forward: 5'-GACTGGTTCCAATTGACAAGC-3' (SEQ ID NO: 5)

20 Reverse: 5'-GCAAATGGCATTCTGACATCC-3' (SEQ ID NO: 6).

Thirty-five cycles of 94°C for 1 minute, 54°C for 1 minute and 72°C for 1 minute were used for PCR in a thermocycler (Model PTC150; MJ Research, Watertown, MA).

EXAMPLE 6

25 Induction of Antibody Expression in Recombinant

P. pastoris Clones

This example illustrates procedure used for inducing antibody expression in recombinant *P. pastoris* clones.

A transformant producing high levels of recombinant
30 antibody was cultured overnight in BMGY broth (1.34% YNB without amino acids or ammonium sulfate, 1.0% glycerol, and 0.4 mg biotin/l) at 30°C for 2 days with shaking at 250 rpm.

The cells were collected by centrifugation and transferred to an inducing medium (1.0% casamino acids,
35 basal medium, trace elements, pH 5.5-6.0, 0.5% methanol and

0.004% biotin). Beginning on the second day and up to fourth day of growth, methanol was added daily to a concentration of 0.5% (v/v), to induce recombinant protein production, and the culture medium and cells were collected separately after low speed centrifugation at 4°C and stored at 20°C. One hundred μ l of a protease inhibitor cocktail (0.35 mg PMSF, 0.31 mg benzamidine, 0.2 mg aprotinin, 0.24 mg pepstain A, 0.2 mg leupeptin, 0.2 mg phenanthroline; Sigma, St. Louis, MO) was added to each induced culture just before harvesting. Slot blots of nitrocellulose membrane containing ten times concentrated supernatants (0.25 ml each) or cell lysates were blocked with nonfat dry milk (10%, w/v, in TBST) for 1 hour, probed with HRP-goat anti-mouse IgG, and developed using chromogenic TMB Microwell Peroxidase Substrate System (Pierce, Rockford, IL).

EXAMPLE 7

Northern Blotting

This example describes conditions used for Northern blotting analysis.

Cells from 5-ml induced cultures of the yeast clones were used for total RNA extraction using a standard method. The cells, harvested by centrifugation at 1000 x g for 10 minutes at 4°C were resuspended in 400 μ l TLS buffer (10 mM Tris-Cl₂ pH 7.4, 1.0% SDS, 100 mM LiCl) and were extracted with TLS buffer-saturated phenol, followed by phenol:chloroform.

Total RNA was ethanol-precipitated, washed with 70% ethanol, air-dried and resuspended in DEPC-treated TLS buffer, and electrophoresed on a 1% agarose gel containing formaldehyde (1.2%) obtained from SEAKEM GTG, Rockland, ME. RNA was transferred onto nylon membrane (Hybond-N) overnight. The membrane was hybridized overnight with α^{32} P-labelled light chain PCR product, followed by washings, first with 2x SSC buffer at 37°C for 15 minutes, repeated

for 20 minutes at room temperature, and finally with 1 x SSC buffer at 37°C for 15 minutes. The membrane was then air-dried and exposed to a Kodak X-Omat film for 24 hours at temperature -80°C.

5

EXAMPLE 8Antibody Expression in Transformed *P. pastoris* Clones

This example describes procedure used for detection of antibody expression in transformed *P. pastoris* clones.

A colony of a high producing transformant was cultured
10 in BMGY broth (1.34% YNB without amino acids ammonium sulfate, 1.0% glycerol, and 0.4 mg biotin/l) overnight at 30°C with shaking at 250 rpm for 2 days. The culture was centrifuged and transferred to an inducing medium made up of 1.0% casamino acids, basal medium, trace elements, pH
15 adjusted with ammonia solution to 5.5-6.0, 0.5% methanol and 0.004% biotin). Cells were allowed to grow further in this medium for 2-4 days at 30°C with shaking at 250 rpm, and 100% methanol was added daily to a concentration of 0.5%.

20 After this period of induction, also described in Example 5, and secretion of protein, cells were harvested by centrifugation at 2500 rpm at 4°C, supernatant was decanted and stored at -20°C until needed. 0.5 ml of each culture medium was dispensed into wells of slot blot
25 apparatus containing nitrocellulose membrane, and vacuum applied until liquid drained out completely. The membrane was blocked with nonfat powdered milk made up in TBST buffer to 10%. After one hour of blocking, the blot was developed with anti-mouse antibody conjugated with horse
30 radish peroxidase (HRP) and developed using TMB (Pierce, Rockford, IL).

Proteins in the culture medium of a positive colony were precipitated with ice-cold acetone concentrated for brief centrifugation (10 minutes) and dried under vacuum.
35 The concentrated proteins were then dissolved in 30 μ l

sample buffer and 10 μ l of each resuspended sample was loaded into wells of a 10% SDS-polyarylamide gel. Gel was stained with coomassie brilliant blue R250 for 30 minutes and destained with acetic acid-methanol-water mixture.

5

EXAMPLE 9Western Immunoblot Analysis

This example describes SDS-PAGE analysis and Western blot analysis.

The culture medium (250 μ l) containing recombinant anti-dioxin antibody was concentrated 25-fold by precipitation with acetone, and 10 μ l of the lysates obtained by dissolving each cell pellet in 30 μ l sample buffer were separately resolved by 10% SDS-PAGE (2 hours at 100 V) and were either stained with Coomassie brilliant blue R250 (Sigma, St. Louis, MO) for 30 minutes and destained using acetic acid/methanol/water (5:25:70), or were electroblotted onto a PVDF membrane (Millipore, Bedford, MA), using Tris-glycine-SDS buffer. The blot was blocked with non-fat milk (5%, w/v, in TBST, pH 8.0), incubated with AP-goat anti-mouse IgG (1:5000, in TBST, pH 8.0) for 1 hour and was washed 4 times in TBST with vigorous shaking. After 25 minutes, it was developed in the dark, with BCIP and NBT, in AP buffer, pH 7.5, at room temperature.

25

EXAMPLE 10ELISA Assay

This example describes functional assay using ELISA, for assaying the hapten-binding activity of the recombinant antibody (Antibodies: A Laboratory Manual, Eds. Harlow, E. and Lane, D., Cold Spring Harbor Laboratory (1988)).

Microplates were coated with 100 μ l well⁻¹ of 50 mM bicarbonate buffer, pH 9.6, 10-50 ng of hapten for 3 hours at 37°C, followed by overnight incubation at 4°C. The plates were then equilibrated at room temperature for 45 minutes, and blocking for non-specific binding was performed for 1

35

hour at 37°C with Tris-buffered saline (TBS; 10 mM Tris-Cl, pH 7.5, 150 mM NaCl) containing 1% (w/v) TBST and 3% BSA.

Plates were washed 3 x with TBST and incubated for 2 hours at room temperature with HRP-goat anti-mouse IgG.

5 The plates were again washed 4 x with TBST, and the HRP activity of the bound antibody was assayed for 10 minutes at 37°C, using the substrates: 0-phenylene diamine (OPD) (30 mg) and H₂O₂ (30 µl), dissolved in 75 ml phosphate-citrate buffer (0.1 M citric acid, 0.2 M Na₂HPO₄, pH 5.0). The

10 reaction was terminated by the addition of 1 M H₂SO₄ (100 µl), and the plates were read at 492 nm, using the UVmax kinetic microplate reader (Molecular Devices Corp., Menlo Park, CA).

All samples were normalized against a sample blank,

15 and read and analyzed using the software package, Softmax (Molecular Devices, Menlo Park, CA).

EXAMPLE 11

Slot-Immunoblot Analysis

This example describes slot-immunoblot analysis of

20 hapten-binding activity of recombinant antibody.

The ability of the recombinant antibody to bind its cognate hapten was assayed using slot-blot approach. The hapten (BSA-dioxin, diluted 1:3,600, to 250 mg/slot⁻¹) was spotted in 5 or 10 µl volumes onto nitrocellulose membrane,

25 using a slot-blot apparatus (Schleicher & Schuell, Keene, NH).

The blot was allowed to air-dry, blocked with non-fat powdered milk (10%, w/v, in Tris-NaCl-Tween 20) at room temperature for 1 hour, and was then incubated with mouse

30 anti-c-myc antibody (diluted 1:5000 in TBST buffer) for 1 hour at room temperature. The blot was then washed 4 x with TBST buffer, incubated with AP-goat anti-mouse IgG (1:5000, in TBST, pH 8.0) for 1 hour at room temperature, again washed with vigorous shaking 5 x over a period of 30

35 minutes, and developed in the dark, with BCIP and NBT, in

AP buffer, pH 7.5 at room temperature.

EXAMPLE 12 .

Kinetics of Recombinant Antibody Synthesis

5 This example illustrates kinetics of recombinant antibody synthesis and secretion.

The kinetics of antibody production and secretion were followed using slot-immunoblots. Single colonies of the clones 11505-1 and 112535-1 and the vector control grown in BMGY broth were induced in MMH medium, and culture media
10 and cells were collected at 12-hour intervals between 0 and 120 hours. Culture media (500 μl slot⁻¹) and cell lysates (10 μl slot⁻¹) were slot-blotted onto a nitrocellulose membrane, blocked and probed with AP-goat anti-mouse IgG as described for the Western immunoblot and in Example 10.

15

WHAT IS CLAIMED:

1. A method for a large-scale production of antigen-specific intact antibody, said method comprising steps:

5 (a) isolating cDNA, mRNA or genomic DNA of genes for antibody light and heavy chains and assembling the antibody genes into expression cassettes containing the cDNA;

(b) preparing a recombinant *P. pastoris* yeast expression vector;

10 (c) constructing a recombinant *P. pastoris* yeast expression plasmid containing the expression cassettes of cDNA of the light and heavy chain genes encoding the antibody;

15 (d) cloning the antibody expression cassettes into the *P. pastoris* expression vector to generate recombinant plasmid;

(e) transforming *Saccharomyces cerevisiae* with the recombinant plasmid by placing said expression cassettes under the control of the AOX1 promoter fused to a *Saccharomyces cerevisiae* α -mating factor signal sequence;

20 (f) amplifying and isolating the recombinant plasmid;

(g) preparing and transforming *P. pastoris* with *Bgl*III, *Not*I, *Sac*I, *Sal*I or *Stu*I-linearized recombinant plasmid replacing the yeast chromosomal AOX1 sequence with AOX1-antibody gene cassettes of the recombinant plasmid;

25 (h) selectively growing the recombinants;

(i) screening yeast transformation colonies for a recombinant antibody expression;

30 (j) analyzing putative positive yeast clones for chromosomal integrates of the expression cassettes of heavy and light chain cDNAs;

(k) confirming the integrity of the DNA insert or junction sequence;

(l) inducing the recombinant antibody expression;

35 (m) confirming the intactness of the expression cassettes inserts with PCR and Northern blot analysis;

(n) detecting the presence of the recombinant antibody by Western blot; and

(o) testing the recombinant antibody for specific antigen-antibody binding.

5

2. The method of claim 1 wherein the antibody genes are assembled into the expression cassettes by subcloning the antibody light and heavy chain cDNA in tandem *EcoRI*-*BglIII/BsmBI* fragments flanked by a *P. pastoris* signal sequence, preceded by a *P. pastoris* promoter at the 5'-terminus and a *P. pastoris* yeast transcription termination sequence at the 3'-terminus.

3. The method of claim 2 wherein the signal sequence is α -factor and wherein the promoter is AOX1-P.

4. The method of claim 3 wherein the yeast expression vector is pPICZ α .

5. The method of claim 4 wherein the yeast expression vector is prepared by restriction digestion with *EcoRI* and *BamHI*.

6. The method of claim 5 wherein the recombinant plasmid is pPICZ α LH.

7. The method of claim 6 wherein the recombinant expression plasmid pPICZ α LH is constructed by cloning the antibody genes expression cassettes into the *P. pastoris* expression vector.

8. The method of claim 7 wherein the replacement of the yeast chromosomal AOX1 with AOX1-antibody gene cassettes is by homologous recombination replacement.

35

9. The method of claim 8 wherein the selective growth of the recombinants is performed on a medium containing zeocin.

5 10. The method of claim 9 wherein the selective growth of the recombinants is performed on a medium containing g418, trimethoprin, or a compound that limits the growth of wild type *P. pastoris*.

10 11. The method of claim 10 wherein the screening of transformed colonies is by colony-immunoblotting.

12. The method of claim 11 wherein the screening is by PCR or by restriction analysis.

15

13. The method of claim 12 wherein the integrity of the DNA inserts or junction sequence is confirmed by nucleotide sequence analysis.

20 14. Intact antigen-specific antibodies produced by *P. pastoris* transformed with mouse, humanized mouse or human immunoglobulin genes, said antibody produced by the process comprising steps:

25 (a) isolating cDNA, mRNA or genomic DNA of genes for antibody light and heavy chains and assembling the antibody genes into expression cassettes containing the cDNA;

(b) preparing a recombinant *P. pastoris* yeast expression vector;

30 (c) constructing a recombinant *P. pastoris* yeast expression plasmid containing the expression cassettes of cDNA of the light and heavy chain genes encoding the antibody;

35 (d) cloning the antibody expression cassettes into the *P. pastoris* expression vector to generate recombinant plasmid;

- (e) transforming *Saccharomyces cerevisiae* with the recombinant plasmid by placing said expression cassettes under the control of the AOX1 promoter fused to a *Saccharomyces cerevisiae* α -mating factor signal sequence;
- 5 (f) amplifying and isolating the recombinant plasmid;
- (g) preparing and transforming *P. pastoris* with *Bgl*III, *Not*I, *Sac*I, *Sal*I or *Stu*I-linearized recombinant plasmid replacing the yeast chromosomal AOX1 sequence with AOX1-antibody gene cassettes of the recombinant plasmid;
- 10 (h) selectively growing the recombinants;
- (i) screening yeast transformation colonies for a recombinant antibody expression;
- (j) analyzing putative positive yeast clones for chromosomal integrates of the expression cassettes of heavy
- 15 and light chain cDNAs;
- (k) confirming the integrity of the DNA insert or junction sequence;
- (l) inducing the recombinant antibody expression;
- (m) confirming the intactness of the expression
- 20 cassettes inserts with PCR and Northern blot analysis;
- (n) detecting the presence of the recombinant antibody by Western blot; and
- (o) testing the recombinant antibody for specific antigen-antibody binding and intactness.

25

15. The antibody of claim 14 wherein the antibody genes are assembled into the expression cassettes by subcloning the antibody light and heavy chain cDNA in tandem *Eco*RI-*Bgl*III/*Bsm*BI fragments flanked by a *P. pastoris* signal sequence, preceded by a *P. pastoris* promoter at the

30 5'-terminus and a *P. pastoris* yeast transcription termination sequence at the 3'-terminus.

16. The antibody of claim 15 produced by *P. pastoris*

35 transformed with human immunoglobulin genes.

17. The antibody of claim 15 produced by *P. pastoris* transformed with humanized mouse immunoglobulin genes.

18. The antibody of claim 15 produced by *P. pastoris*
5 transformed with mammalian or mouse immunoglobulin genes.

19. A recombinant *P. pastoris* yeast expression vector containing dual expression cassettes, each carrying a cDNA copy of immunoglobulin light and heavy chain.
10

20. An expression system comprising *P. pastoris* transformed with antibody genes for production of a recombinant antigen-specific intact antibody.

15 21. *P. pastoris* yeast transformed with expression cassettes carrying a cDNA copy of immunoglobulin heavy and light chain suitable for large-scale production of intact antibodies.

FIG. 4

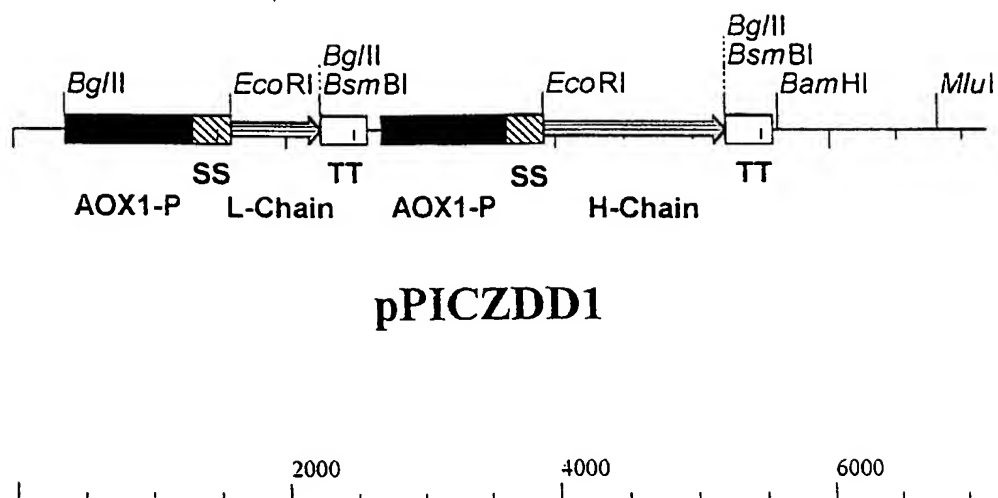
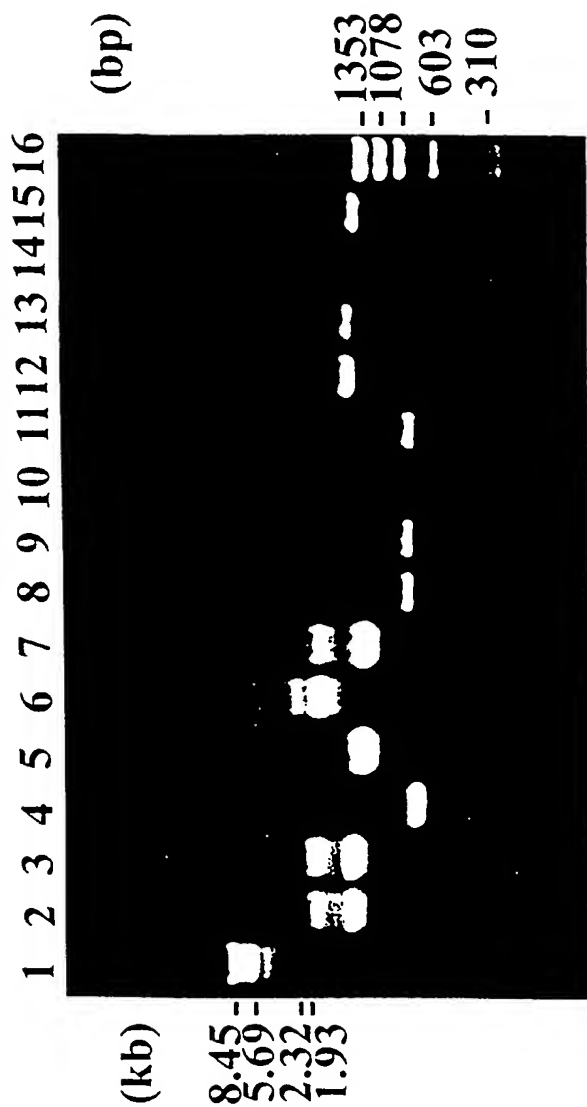
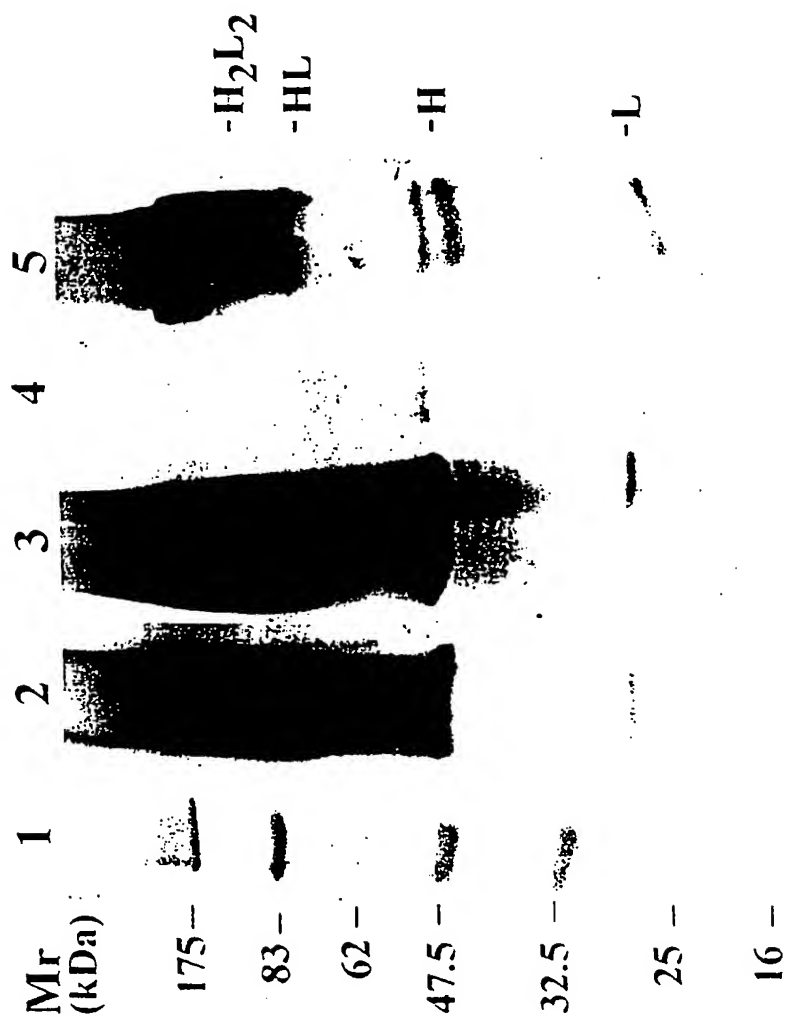


FIG. 2



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FIG. 4



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FIG. 5

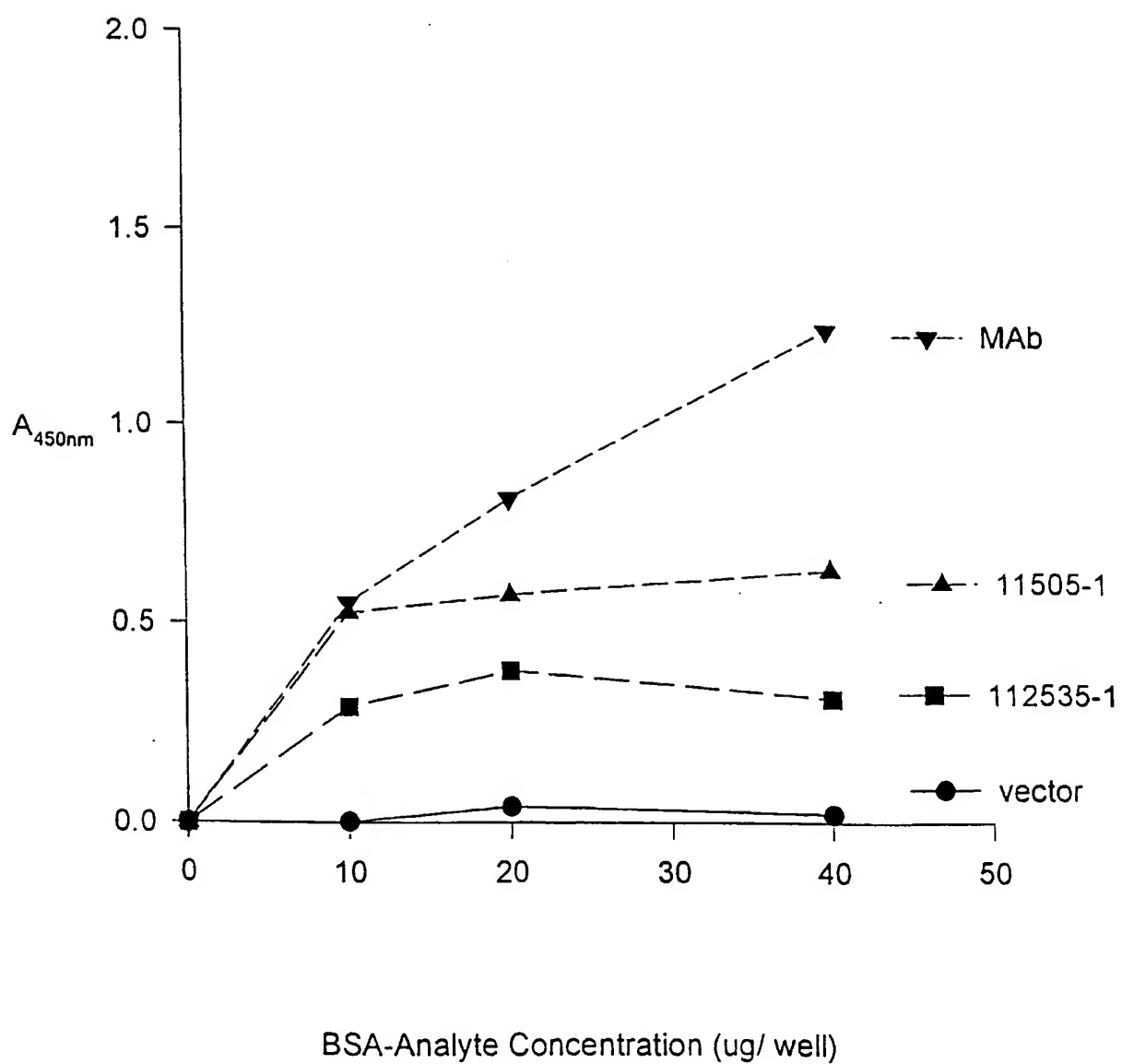
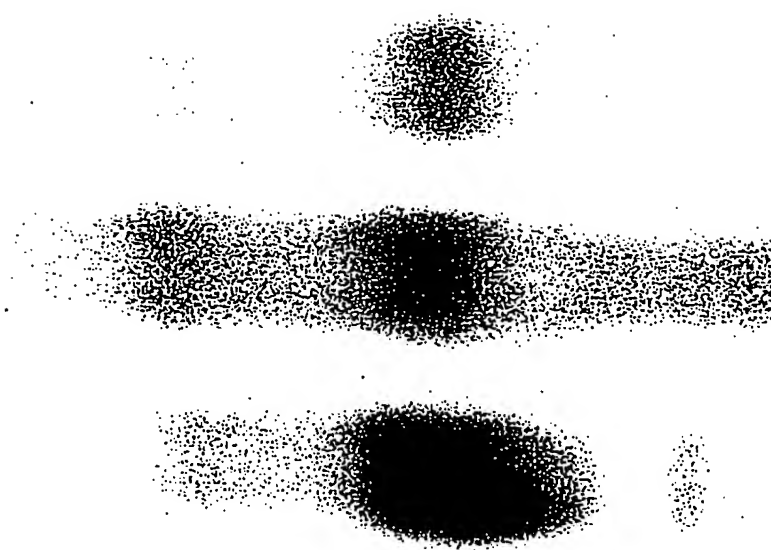


FIG. 3

1 2 3 4

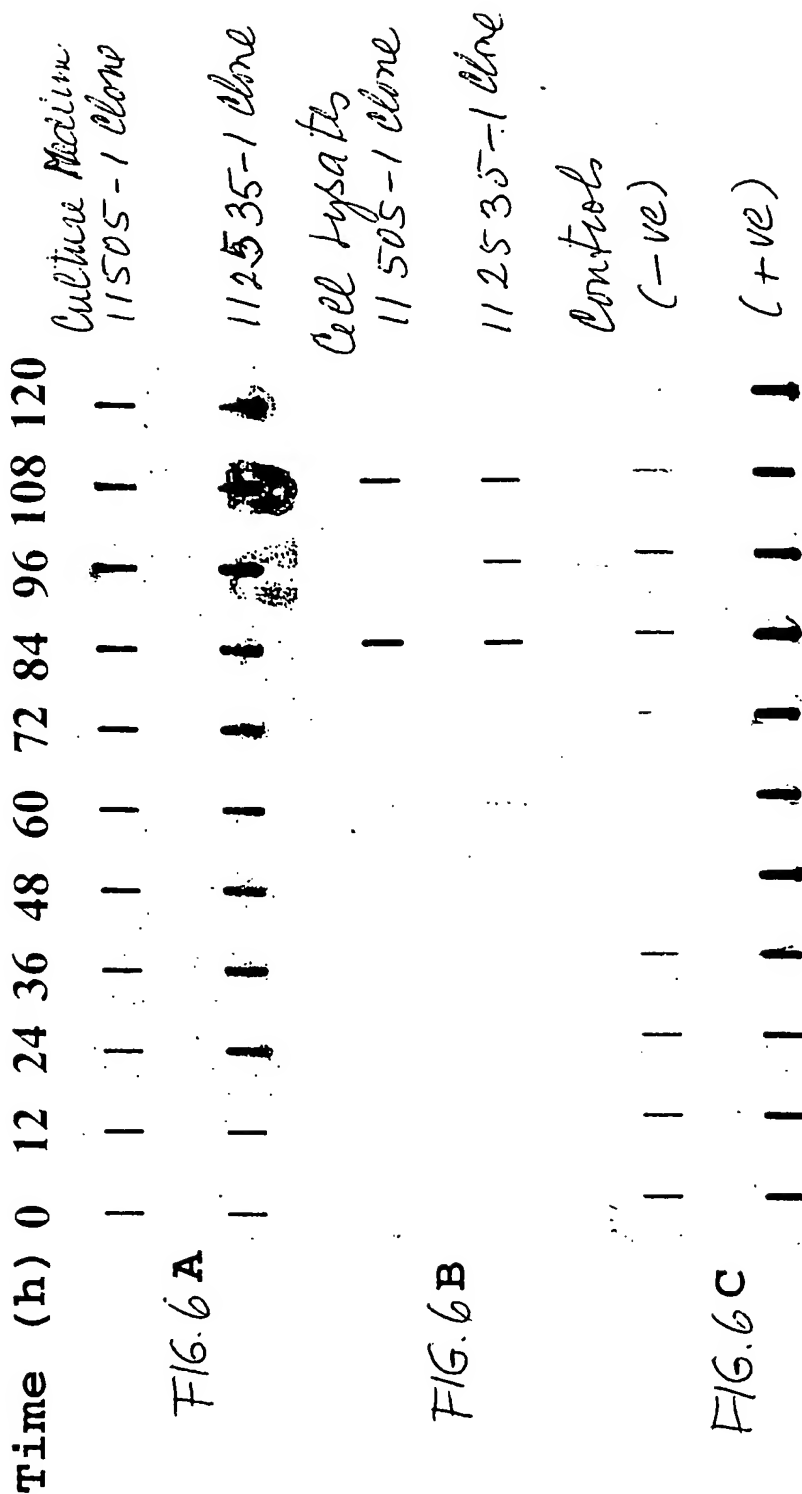
- 2.0 kb

- 1.35 kb



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FIG. 6



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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/24677

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : Please See Extra Sheet.

US CL : Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/69.1, 69.9, 255.1; 530/387.1, 387.3; 536/23.1, 23.53; 436/501; 424/130.1, 133.1, 141.1, 142.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	HORWITZ et al. Secretion of functional antibody and Fab fragment from yeast cells. Natl. Acad. Sci., USA. November 1988, Vol. 85, pages 8678-8682, especially page 8678 and Figure 2.	14, 16-21
--		1-13, 15
Y	US 5,102,789 A (SIEGEL et al) 07 April 1992, col. 3, lines 50-60; col. 7, lines 42-49; col. 10, lines 9-15 and 55-62; col. 11, lines 24-27.	1-21
Y	US 5,750,372 A (SAKAI et al) 12 May 1998, abstract; col. 4, lines 41-52; col. 6, lines 8-10.	1-21
X	US 5,783,431 A (PETERSON et al) 21 July 1998, col. 59, lines 45-47.	1, 10

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
B earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*A* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

15 JANUARY 2000

Date of mailing of the international search report

04 FEB 2000

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/24677

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 4,816,397 A (BOSS et al) 28 March 1989, abstract; col. 6, lines 1-4; col. 5, lines 37-56, cols. 22-23.	1-21
Y	Invitrogen Catalog. "anonymous", January 1998. Yeast Expression.	1-9

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/24677

A. CLASSIFICATION OF SUBJECT MATTER:

IPC (7):

C12N 15/00, 15/12, 15/13, 15/81; C07K 16/00; G01N 33/50; A61K 39/395

A. CLASSIFICATION OF SUBJECT MATTER:

US CL :

435/69.1, 69.9, 255.1; 530/387.1, 387.3; 536/23.1, 23.53; 436/501; 424/130.1, 133.1, 141.1, 142.1

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

CAPLUS, MEDLINE, USPATFULL, WEST

search terms: antibody, yeast A)X1 promoter, yeast expression vector, pPICZ, zeocin, alpha-mating factor, P. pastoris, S. cerevisiae, g418, humanized, murine antibody, chimeric